REPORT DOCUMENTATION PAGE

Form Approved OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.

1. REPORT DATE (DD-MM-YYYY)	2. REPORT TYPE	3. DATES COVERED (From - To)
16 Nov 2012	FINAL	1 AUG 2010-31 July 2012
I. TITLE AND SUBTITLE		5a. CONTRACT NUMBER
Therapeutic Targeting of P2X7 after TBI		N/A
		5b. GRANT NUMBER
		HU0001-10-1-TS11
		5c. PROGRAM ELEMENT NUMBER
		N/A
S. AUTHOR(S)		5d. PROJECT NUMBER
Kimbler, Donald, PhD, RN, I	MAJ, AN, USA	N10-P10
		5e. TASK NUMBER
		N/A
		5f. WORK UNIT NUMBER
		N/A
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)	8. PERFORMING ORGANIZATION REPORT
Georgia Health Sciences Un.		NUMBER
1120 15 th St, CJ 3301		N/A
Augusta, GA 30912		
. SPONSORING / MONITORING AGENCY	NAME(S) AND ADDRESS(ES)	10. SPONSOR/MONITOR'S ACRONYM(S)
TriService Nursing Research	` ,	TSNRP
Program, 4301 Jones Bridge		
Bethesda, MD 20814		11. SPONSOR/MONITOR'S REPORT
2001100000, 1112 20011		NUMBER(S)
		N10-P10
2 DISTRIBUTION / AVAIL ARILITY STATE	MENT	1.10 110

12. DISTRIBUTION / AVAILABILITY STATEMENT

Approved for public release; distribution unlimited

13. SUPPLEMENTARY NOTES

N/A

14. ABSTRACT

Traumatic brain injury (TBI) is a leading cause of death and disability worldwide. Cerebral edema, the abnormal accumulation of fluid within the brain parenchyma, contributes to elevated intracranial pressure (ICP) and is a common life-threatening neurological complication following TBI. Unfortunately, neurosurgical approaches to alleviate increased ICP remain controversial and medical therapies are lacking due in part to the absence of viable drug targets. In the present study, genetic inhibition (P2X7-/- mice) of the purinergic P2x7 receptor attenuated the expression of the pro-inflammatory cytokine, interleukin-1β (IL-1β) and reduced cerebral edema following controlled cortical impact, as compared to wild-type mice. Similarly, brilliant blue G (BBG), a clinically non-toxic P2X7 inhibitor, inhibited IL-1β expression, limited edemic development, and improved neurobehavioral outcomes after TBI. The beneficial effects of BBG followed either prophylactic administration via the drinking water for one week prior to injury or via an intravenous bolus administration up to four hours after TBI, suggesting a clinically-implementable therapeutic window. Notably, P2X7 localized within astrocytic end feet and administration of BBG decreased the expression of glial fibrillary acidic protein (GFAP), a reactive astrocyte marker, and attenuated the expression of aquaporin-4 (AQP4), an astrocytic water channel that promotes cellular edema. Together, these data implicate P2X7 as a novel therapeutic target to prevent secondary neurological injury after TBI, a finding that warrants further investigation.

15. SUBJECT TERMS

Traumatic brain injury (TBI), elevated intracranial pressure (ICP), secondary neurological injury, military deployment

16. SECURITY CLASS	SIFICATION OF:		17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON Debra Esty
a. REPORT UNCLASSIFIED	b. ABSTRACT UNCLASSIFIED	c. THIS PAGE UNCLASSIFIED	טט	41	19b. TELEPHONE NUMBER (include area code) 301-319-0596

TriService Nursing Research Program Final Report Cover Page TriService Nursing Research Program

Inservice Nursing Research Program
4301 Jones Bridge Road Bethesda MD 20814
HU0001-10-1-TS11
N10-P10
Therapeutic Targeting of P2X7 after TBI
1 August 2010 - 31 July 2012
Georgia Health Sciences University
1120 15 th St, CJ 3301, Augusta, GA 30912
Date
Date

Table of Contents

Abstract	3
TSNRP Research Priorities that Study or Project Addresses	4
Progress Towards Achievement of Specific Aims of the Study or Project	5
Specific Aim 1 Specific Aim 2 Specific Aim 3 Specific Aim 4	5 8 13 15
Discussion	16
References Cited	23
Summary of Dissemination	39
Reportable Outcomes	40

Kimbler, Donald E

N10-P10

Abstract

Traumatic brain injury (TBI) is a leading cause of death and disability worldwide. Cerebral edema, the abnormal accumulation of fluid within the brain parenchyma, contributes to elevated intracranial pressure (ICP) and is a common life-threatening neurological complication following TBI. Unfortunately, neurosurgical approaches to alleviate increased ICP remain controversial and medical therapies are lacking due in part to the absence of viable drug targets. In the present study, genetic inhibition (P2X7-/mice) of the purinergic P2x7 receptor attenuated the expression of the pro-inflammatory cytokine. interleukin-1ß (IL-1ß) and reduced cerebral edema following controlled cortical impact, as compared to wild-type mice. Similarly, brilliant blue G (BBG), a clinically non-toxic P2X7 inhibitor, inhibited IL-1β expression, limited edemic development, and improved neurobehavioral outcomes after TBI. The beneficial effects of BBG followed either prophylactic administration via the drinking water for one week prior to injury or via an intravenous bolus administration up to four hours after TBI, suggesting a clinicallyimplementable therapeutic window. Notably, P2X7 localized within astrocytic end feet and administration of BBG decreased the expression of glial fibrillary acidic protein (GFAP), a reactive astrocyte marker, and attenuated the expression of aquaporin-4 (AQP4), an astrocytic water channel that promotes cellular edema. Together, these data implicate P2X7 as a novel therapeutic target to prevent secondary neurological injury after TBI, a finding that warrants further investigation

3

TSNRP Research Priorities that Study or Project Addresses

Primary Priority

1 I IIIIai y 1 I Ioi ity	
Force Health Protection:	☐ Fit and ready force ☐ Deploy with and care for the warrior ☐ Care for all entrusted to our care
Nursing Competencies and Practice:	☐ Patient outcomes ☐ Quality and safety ☐ Translate research into practice/evidence-based practice ☐ Clinical excellence ☐ Knowledge management ☐ Education and training
Leadership, Ethics, and Mentoring:	 ☐ Health policy ☐ Recruitment and retention ☐ Preparing tomorrow's leaders ☐ Care of the caregiver
Other:	☐ Translating Knowledge & Research Findings into Practice in a Military Context

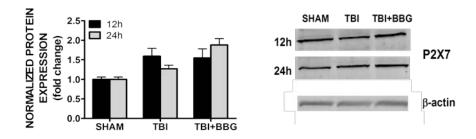
Progress towards Achievement of Specific Aims of the Study or Project The overall objective of this research was to elucidate molecular and cellular mechanisms that promote cerebral edema, which may aid in the development of novel therapeutics to limit neurological dysfunction and reduce the incidence of neuropsychiatric sequelae following TBI. The purinergic receptor P2X7 has been implicated in the processing and or release of interleukin 1 beta (IL-1 β), the prototypical, pro-inflammatory cytokine. A low affinity ATP activated receptor, P2X7 plays a role in the innate immunity and upon activation causes trans-membrane ion fluxes and formation of membrane pores as well as its role in the production and release of IL-1 β . Others have shown a positive correlative role between IL-1 β and the development of cerebral edema after brain injury. To this end we proposed a mechanistic hypothesis to explain the etiology of cerebral edema, which if shown, could potentially lead to new therapies in TBI.

Specific Aim 1: To establish the cellular localization and temporal pattern of P2X7 expression following TBI.

Hypothesis. The purinergic receptor, P2X7, is activated following TBI.

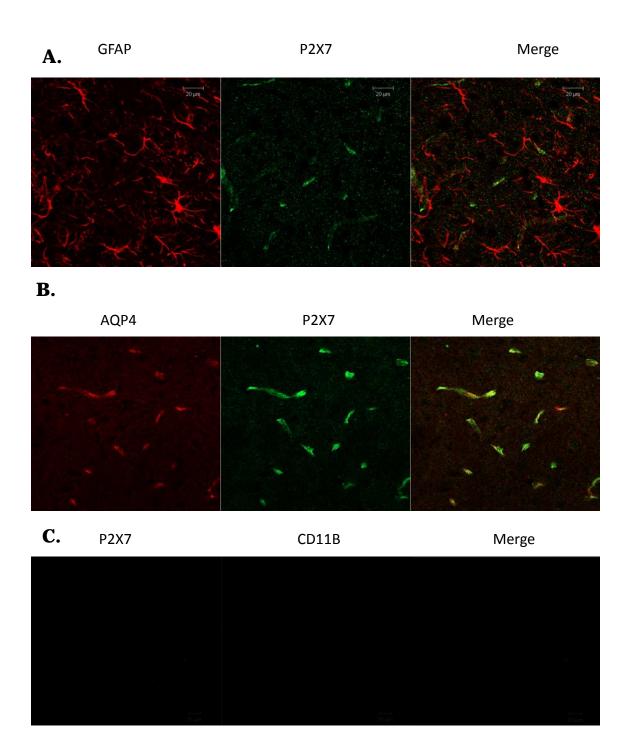
Rationale. The P2X7 receptor is reportedly distributed on cells of the immune system including macrophages, monocytes, lymphocytes, etc; additionally, the receptor is found in glia cells of the central and peripheral nervous system as well as spinal cord neurons (Coddou, Yan, Obsil, Huidobro-Toro, & Stojilkovic, 2011; Wang, et al., 2004). Cellular localization of the P2X7 receptor has yet to be clearly defined after TBI; therefore these studies will show, for the first time, the temporal pattern and cellular localization of P2X7 in the brain following TBI.

The expression of P2X7, the presumed cellular target of BBG action, was next assessed within the brain. P2X7 was basally expressed within the cerebral cortex, as demonstrated by Western blotting; however, expression was not increased following TBI, as compared to sham-operated mice (**Figure 1**).



<u>Figure 1: Western blot analysis of P2X7 receptor after TBI</u>. Representative Western blots (top panel) of P2X7 in the cerebral cortex of mice following sham injury, TBI, or TBI + 50 mg/kg BBG. Tissue was collected at 12h or 24h after TBI. Blots were normalized to β-actin to control for equal protein loading between lanes. Data are representative of six mice/group. Densitometric analysis of Western blots (bottom panel) is presented as P2x7 expression following normalization to β-actin

Immunohistochemical analysis revealed that P2X7 strongly co-localized with the astrocytic endfoot marker, aquaporin-4 (AQP4) (**Figure 2b**) whereas dual labeling was not observed with the microglial marker, CD11b (**Figure 2c**) or the astrocytic cell body marker, glial fibrillary acidic protein (GFAP) (**Figure 2a**). Furthermore, no localization was observed between P2X7 and the neuron-specific marker, NeuN (data not shown). Together, these data implicate astrocytes as a key mediator of the biological actions of P2X7 and as a possible cellular target of BBG after TBI.



Specific Aim 2: To establish whether inhibition of P2X7 decreases cerebral edema following TBI.

Hypothesis. Antagonism of the purinergic receptor, P2X7, will reduce cerebral edema after TBI.

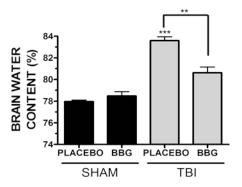
Rationale. ATP is generally contained to intracellular compartments and is not commonly found in the external milieu. After injury, ATP is released into the external environment where it can act upon the low affinity purinergic receptor P2X7. The P2X7 receptor has been implicated in neuropathic pain and inflammatory responses throughout the body. Upon P2X7 well known to open channels that are that are permeable to both mono and divalent cations, altering concentrations that can fluxes in water content. Work by Nedergaard and colleagues showed that doses of 10mg/kg and 50mg/kg of BBG, a specific P2X7 antagonist, showed improvements after spinal cord injury. The proper dose and effect of BBG in the brain remains largely unexplored; therefore, these studies will implicate P2x7 in edema development after TBI. If successful, these studies will also identify a novel therapeutic use for brilliant blue G (BBG), a clinically- safe P2X7 antagonist, amendable to implementation on the battlefield (Wang, et al., 2004).

BBG reduces post-traumatic cerebral edema with an extended therapeutic window

Brain water content, a sensitive measure of cerebral edema, was significantly increased

within the ipsilateral cortex at 24h post-TBI (83.6 \pm 0.4% brain water content after TBI vs. 77.9 \pm 0.2% in sham, p<0.001 vs. sham) (**Figure 3**). A single, intravenous injection of 50 mg/kg BBG at 15 minutes

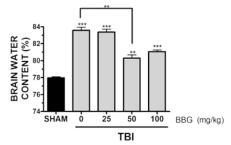
prior to injury attenuated brain water content after TBI ($80.6 \pm 0.5\%$; p<0.01 vs. TBI) whereas administration of 25 mg/kg BBG did not significantly reduce edema ($83.3\% \pm 0.3\%$; not significantly different from TBI). Notably, the ability of 100 mg/kg BBG to reduce edema was not significantly different from administration of 50



<u>Figure 3: Antagonism of P2X7 reduces cerebral</u> <u>edema after TBI.</u>

A single intravenous bolus of 50 mg/kg BBG provided 15 minutes prior to TBI, significantly reduced the development of cerebral edema at 24h post-TBI, as measured by brain water content. Comparisons within each hemisphere between different treatments groups were done using a one-way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01, ***p<0.001 vs. the ipsilateral hemisphere in sham-operated mice). No significant differences in cerebral edema were observed between groups in the contralateral hemisphere. Data are represented as the mean \pm SEM from 5-6 mice/group

mg/kg (81.0 \pm 0.2%; p<0.001 vs. TBI), suggesting 50 mg/kg was the lowest efficacious dose to limit edemic development after TBI (**Figure 4**). For all studies, brain water content



<u>Figure 4: Dose determination of BBG for antagonism of P2X7 after</u>

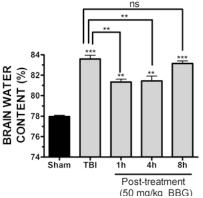
TBI. A single intravenous bolus of 50-100 mg/kg BBG administered 0.5h after TBI significantly reduced cerebral edema at 24h post-TBI. . Comparisons within each hemisphere between different treatments groups were done using a one-way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01, ***p<0.001 vs. the ipsilateral hemisphere in sham-operated mice). No significant differences in cerebral edema were observed between groups in the contralateral hemisphere. Data are represented as the mean ± SEM from 5-6 mice/group

within the contralateral (uninjured) cortices did not significantly differ between any of the treatment groups (data not shown). Furthermore, administration of BBG alone (50 mg/kg, i.v., 15 minute pre-treatment) did not significantly change brain water content, as compared to placebo-treated, sham-operated mice (**Figure 3**), suggesting an injury-specific reduction in edema.

The therapeutic window whereby BBG reduced edemic development was next established. A 1h post-treatment with 50 mg/kg significantly reduce cerebral edema (81.3 \pm 0.2%, p<0.05 vs. TBI) to a similar extent as pre-

 \pm 0.2%, p<0.05 vs. TBI) to a similar extent as pretreatment (**Figure 5**; see **Figure 3** for comparison). Similarly, a 4h post-treatment effectively attenuated post-traumatic edema (81.4 \pm 0.4%, p<0.05 vs. TBI, no significantly different from 1h post-treatment). In contrast, 8h post-treatment with 50 mg/kg was ineffective at reducing edema, as compared to TBI (83.2% \pm 0.2%), suggesting a 4h post-injury therapeutic window.

We next determined whether prophylactic administration of BBG may reduce edema. Oral administration of 25 mg/mL BBG via the drinking



<u>Figure 5: Determination of BBG time of treatment after TBI.</u>

Administration of a single intravenous bolus of 50 mg/kg BBG reduced cerebral edema when administered 1h or 4h after injury. This effect was lost if post-treatment was delayed beyond 8h from the time of injury. Comparisons within each hemisphere between different treatments groups were done using a one-way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01, ***p<0.001 vs. the ipsilateral hemisphere in sham-operated mice). No significant differences in cerebral edema were observed between groups in the contralateral hemisphere. Data are represented as the mean ± SEM from 5-6 mice/group

water for one week prior to injury effectively decreased brain edema after TBI ($80.9 \pm 0.2\%$, p<0.01 vs. TBI) (**Figure 6**). In contrast, 10 mg/mL BBG via the drinking water did not significantly reduce edema, as compared to mice receiving water containing only placebo. As a whole, either prophylactic oral administration or post-injury intravenous administration of BBG effective attenuates brain edema after TBI.

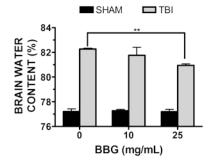


Figure 6: Prophylactic treatment with BBG reduces cerebral edema after TBI. Prophylactic treatment with BBG in the drinking water for 7 days reduced edema at 24h post-TBI at a concentration of 25 mg/ml but not 10mg/ml. Comparisons within each hemisphere between different treatments groups were done using a one-way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01, ***p<0.001 vs. the ipsilateral hemisphere in sham-operated mice). No significant differences in cerebral edema were observed between groups in the contralateral hemisphere. Data are represented as the mean \pm SEM from 5-6 mice/group

Brain expression of P2X7 after TBI

Peripheral administration of BBG reduced brain edema, although the potential tissue and cellular targets of BBG remained unclear. Intravenous administration of 50 mg/kg BBG produced a transient deep blue color over the first 24h within the eyes, nose, ears, and paws (**Figure 7a**), demonstrating wide peripheral distribution throughout the circulatory system. No trace of blue color was observed by 72h post-administration. Similarly, oral administration of BBG for one week via the drinking water also produced a faint blue hue in the paws and eyes, albeit to a far lesser extent, as compared to intravenous administration. Consistent with the observed blue appearance, serum levels of BBG reached $383 \pm 33.3 \mu M$ and 1.73 ± 0.07 mM following intravenous administration of 50 mg/kg and 100 mg/kg, respectively. Whether BBG acted peripherally or crossed the blood-brain barrier to directly affect the brain after TBI remained unclear. Consistent with a potential direct effect, the brains of mice administered BBG appeared greyish-blue, with blue color observed within the cerebral vasculature and brain tissue. Most notably, the contused cortex exhibited a distinct blue color (**Figure 7b**), suggesting BBG can enter the brain and preferentially accumulates at high levels around damaged tissue, presumably where the blood-brain barrier is disrupted.

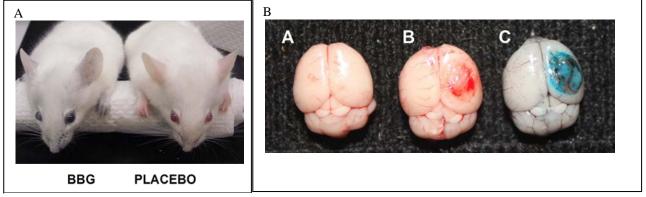


Figure 7: Distribution of BBG after TBI. (A) Photograph of representative mice following an intravenous administration of placebo (right) and a BBG (50 mg/kg; left). Note the blue appearance in the skin, eyes, ears, paws and tail. **(B)** BBG accumulates in the contused cortex after TBI. Photographs of brains taken from a sham-operated mouse administered placebo (left panel), a mouse administered placebo at 0.5h after TBI (middle panel), or a mouse administered 50 mg/kg BBG via the tail vein at 0.5h post-TBI.

P2X7-/- mice exhibit reduced cerebral edema after TBI

BBG is a highly selective inhibitor of P2X7; however, pharmacological agents often exhibit "off-target" or non-specific effects. To validate P2X7 as a potential therapeutic target to reduce brain edema, we next performed studies in P2X7-/- mice. Consistent with data collected after BBG administration, P2X7-/- mice exhibited a significant reduction in

brain water content, as compared to wild-type mice, following TBI ($81.0 \pm 0.4\%$ in P2X7-/- vs. $83.7 \pm 0.3\%$ in wild-type; p<0.01) (**Figure 8**). These findings were supported by the measurement of edemic volume in living mice using MRI. P2X7-/- mice exhibited a 36% reduced in edemic volume after TBI, as compared to wild-type mice (14.4 ± 0.7 mm³ in wild-type mice vs. 9.2 ± 1.5 mm³ in P2X7-/- mice; p<0.01 vs. wild-type) (**Figure 9**). Brain water content was not

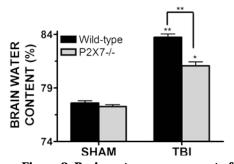
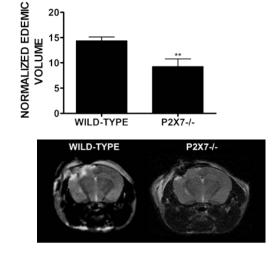


Figure 8: Brain water measurement of genetic inhibition of P2X7 after TBI.
P2X7-/- mice exhibited a significant reduction in brain water content, as compared to wild-type mice, when assessed at 24h post-TBI.
Comparisons within each hemisphere between different treatments groups were done using a one-way ANOVA followed by Dunnett's post-hoc test (* p<0.05 vs. the ipsilateral hemisphere in sham-operated mice). No significant differences in cerebral edema were observed between groups in the contralateral hemisphere.

significantly different either in sham-operated mice (**Figure 9**) or in the contralateral hemisphere of wild-type or P2X7-/- mice.

Figure 9: Genetic



inhibition of P2X7 <u>attenuates cerebral edema</u> after TBI as measured by MRI. P2X7-/- mice displayed attenuated cerebral edema, as compared to wild-type mice, when assessed by MRI. The top panels depict a representative wild-type and a P2X7-/- mouse imaged at 24h post-TBI. Bottom panels represent the mean edemic volume of mice imaged by MRI. Data are represented as the mean \pm SEM from six mice/group and were analyzed using a t-test (p<0.01 vs. wild-type).

Specific Aim 3: To determine whether P2x7 inhibition reduces IL1-1 β production and release following TBI.

Hypothesis. Inhibition of the P2X7 receptor will decrease the production of IL-1 β following TBI

Rationale. The brain has long been thought to be immuno-privileged. Recent work by numerous laboratories has shown that the innate immune system functions similarly if not identically in the brain as in the periphery. The purinergic receptor P2X7, originally described in cells of hematopoetic origin to include microglia, is known to not only allow the bidirectional flow of cations but to have an important role in the release of proinflammatory cytokines such as IL-1 β . IL-1 β is an important mediator in chronic pain, inflammation and neurodegeneration and can affect neuronal cell death after injuries such as TBI. P2X7 activity has been reported to have a role in the pathology of disease processes such as depression by regulating the release of the proinflammatory cytokine IL-1 β . Ito et al (1996) describes a positive correlation between Il-1 β in the CSF, cerebral edema and negative outcomes associated with TBI. This provides the theoretical link between the immune system and neurologic injuries providing the foundation for the studies of this aim. Therefore, these studies will implicate P2X7 activation in the production and release of the pro- inflammatory cytokine IL-1 β after TBI, suggesting an innate immune component of the



B-ACTIN

inflammatory pathway of TBI and cerebral edema

(Skaper, et al., 2010;

Wang, et al., 2004).

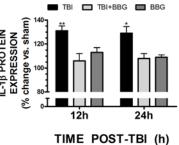


Figure 10: Western blot analysis of IL-1β after inhibition of P2X7 single intravenous bolus of 50-100 mg/kg BBG administered 0.5h after TBI significantly reduced peri-contusional IL-1 β expression, as assessed by **(B)** EIA and by (A) Western blotting at 12h or 24h post-injury. In panel A,data are represented as IL-1 β expression as a % of In panel B. sham expression levels. data was normalized to β -actin to control for equal protein loading between lanes. Data are representative Data were analyzed with mice/group. One-Way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01 vs. sham operated mice).

P2X7 mediates glial reactivity after TBI

IL-1 β induces reactive astrogliosis after TBI; therefore, the ability of BBG to attenuate the expression of GFAP, a hallmark of gliosis, was next assessed. GFAP expression was significantly increased by 299.7 \pm 72.2% within the peri-contusional cortex (p<0.05 vs. sham) and 222.0 \pm 28.6% (p<0.01 vs. sham) of sham-operated mice at 12h and 24h post-

TBI, respectively (**Figure 11**). Post-treatment with 50 mg/kg BBG reduced GFAP expression to $216.1 \pm 88.6\%$ (not significantly different from either sham or TBI) and $145.3 \pm 15.2\%$ (p<0.05 vs. TBI, not significantly different from sham) of expression levels in sham-operated mice at 12h and 24h, respectively.

Consistent with the inhibitory effect of BBG on posttraumatic cerebral edema and glial reactivity, BBG attenuated the expression of the astrocytic water channel, AQP4, after TBI. AQP4 protein expression was increased

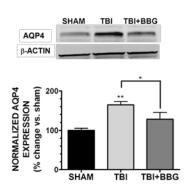


Figure 12: Western blot analysis of AQP4 expression after P2X7 antagonism. Representative Western blot (top panel) of AQP4 in the cerebral cortex of mice at 12h following sham injury, TBI, or TBI + 50 mg/kg BBG. Densitometric analysis of Western blots (bottom panels) is presented as AQP4 expression following normalization to β -actin, which was used to control for equal protein loading. Data (mean \pm SEM) are representative of six mice/group from three independent experiments (n=3/group in each experiment) and are expressed as % change vs. sham. Data were analyzed by One-Way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01 vs. sham operated mice).

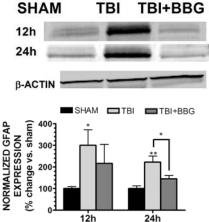


Figure 11: BBG attenuates glial activation as measured by GFAP expression. Representative Western blot (top panel) of cortical GFAP expression taken at 12h or 24h after sham injury, TBI, or TBI + 50 mg/kg BBG. Data (mean ± SEM) are representative of six mice/group from three independent experiments (n=3/group in each experiment) and are expressed as % change vs. sham. Data were analyzed by One-Way ANOVA followed by Dunnett's posthoc test (* p<0.05, ** p<0.01 vs. sham operated mice).

within the pericontusional

context at 12h (1.7 \pm 0.1 fold increase; p<0.01 vs. sham) and at 24h (1.5 \pm 0.1 fold increase; p<0.05 vs. sham)

after TBI. Intravenous administration of 50 mg/kg BBG at 0.5h post-injury attenuated the

TBI-induced increases in AQP4 expression $(1.3 \pm 0.2 \text{ and } 1.1 \pm 0.1 \text{ fold increase vs. sham at } 12\text{h}$ and 24h, respectively; p<0.05 vs. TBI, not significant different from sham) (**Figure 12**).

Specific Aim 4: To determine whether inhibition of P2X7 improves neurologic outcomes following TBI

Hypothesis. *Inhibition of the P2X7 receptor after TBI will improve neurologic outcomes* **Rationale.** The brain has long been thought to be immuno-privileged. Recent work by numerous laboratories has shown that the innate immune system functions similarly if not identically in the brain as in the periphery. The purinergic receptor P2X7, originally described in cells of hematopoetic origin to include microglia, is known to not only allow the bidirectional flow of cations but to have an important role in the release of proinflammatory cytokines such as IL-1 β . IL-1 β is an important mediator in chronic pain, inflammation and neurodegeneration and can affect neuronal cell death after injuries such as TBI. P2X7 activity has been reported to have a role in the pathology of disease processes such as depression by regulating the release of the proinflammatory cytokine IL-1 β . P2X7 inhibition has been reported to improve outcomes after both spinal cord injury and stroke(Arbeloa, Perez-Samartin, Gottlieb, & Matute, 2011; Skaper, et al., 2010; Wang, et al., 2004). Therefore, we propose that these studies will show for the first time that inhibition of P2X7 will improve neurobehavioral outcomes after TBI.

BBG improves neurobehavioral outcomes after TBI

Depression and anxiety disorders are common psychiatric co-morbidities after a TBI. Thus, the ability of BBG to reduce neuropsychiatric dysfunction was next explored. A significant increase in open-field hyperlocomotion (total number of squares entered) was observed following TBI (p<0.01 vs. sham) (**Figure 13**). Administration of 50 mg/kg BBG

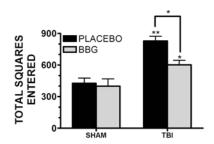


Figure 13: BBG improves open field hyperlocomotion after TBI. Post-injury administration of 50 mg/kg BBG significantly attenuated post-traumatic hyperlocomotion following TBI in the open field test and (B) time to first immobility in the forced swim test, a sensitive estimate of depressive like behavior, as compared to placebo-treated mice. Data are expressed as the mean ± SEM from 10-12 mice/group and were compared by One-Way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01 vs. sham operated mice).

partially resolved the increase in post-traumatic hyperlocomotion by $\sim 50\%$ (p<0.05 vs. sham and TBI). In contrast, BBG administration had no significant effect on basal activity in sham-operated mice.

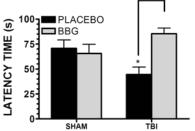
Following TBI, mice exhibited a reduced time to latency to develop behavioral despair, a measure of depression, using the forced swim test. Shamoperated mice displayed a latency of 70.8 ± 8.3 s whereas TBI reduced this time to 44.5 ± 7.4 s (p<0.05

vs. sham). Post-injury administration of 50 mg/kg BBG significantly increased the latency

time to $85.4 \pm 5.5s$ (p<0.01 vs. TBI, not significantly different from sham). (**Figure 14**). Notably, BBG administration did not significantly change the latency time in sham-operated mice, suggesting an injury specific effect.

DISCUSSION

Preventative measures reduce the incidence and/or severity of TBI, yet one-third of hospitalized TBI patients die from injuries that are secondary to the initial trauma.



BBG Figure 14: <u>improves</u> depressive effects after TBI. Postinjury administration of 50 mg/kg BBG significantly increased time to first immobility in the forced swim test, a sensitive estimate of depressive like behavior, as compared to placebotreated mice. Data are expressed as the mean ± SEM from 10-12 mice/group and were compared by One-Way ANOVA followed by Dunnett's post-hoc test (* p<0.05, ** p<0.01 vs. sham operated

The development of post-traumatic edema promotes clinical deterioration and worsens long-term outcomes, at least in part, by limiting cerebral perfusion, by increasing brain herniation, and by increasing the manifestation of neuropsychiatric impairments such as headaches, anxiety, depression, sleep disturbances, and appetite loss (H. S. Levin, et al., 1991; Rogers & Read, 2007; Saul & Ducker, 1982a; Whelan-Goodinson, Ponsford,

Johnston, & Grant, 2009). Thus, elucidation of the cellular mechanisms of neurological injury may permit the development of efficacious therapeutics to improve patient outcomes after TBI.

In the present study, genetic (P2X7-/-) or pharmacological (BBG) inhibition of P2X7 reduced secondary brain injury and improved functional outcomes after a moderate TBI in mice. BBG, a FDA-approved, water soluble, structural and functional analogue of FD&C blue dye No. 1 (also called Brilliant blue FCF or E133), is a widely used food additive and coloring agent that exhibits no toxicity at doses up to 1g/kg/d in humans (Register, 2006). Herein, BBG reduced peri-contusional IL-1β, limited AQP4 expression, attenuated edemic development, and improved neurobehavioral outcomes. These beneficial effects were observed whether BBG was intravenously administered as a single bolus up to four hours after injury or chronically administered via the drinking water. Thus, clinically safe doses of BBG may reduce neurological injury after TBI, either via a clinically-implementable postinjury temporal window or via prophylactic administration.

Cellular edema is the predominant form of edema during the acute and sub-acute phase after TBI (Bullock, Maxwell, Graham, Teasdale, & Adams, 1991; J. Ito, et al., 1996). Astrocytic swelling, a characteristic feature of cellular edema, commenced within the first hours after head trauma in humans (Bullock, et al., 1991; J. Ito, et al., 1996) and glial activation temporally paralleled edemic development in pre-clinical models of TBI (Dietrich, et al., 1999; Hinkle, et al., 1997). Furthermore, increased serum and CSF levels of the activated astrocyte markers, S100 β and GFAP, directly correlated with patient outcomes after TBI (Hayakata, et al., 2004; Pelinka, Kroepfl, Leixnering, et al., 2004; Pelinka, Kroepfl, Schmidhammer, et al., 2004), supporting a possible role for astrocytes in the genesis of secondary neurovascular injury; however, controversy remains as to whether

astrocytes exert beneficial and/or detrimental functions after brain injury (Laird, Vender, & Dhandapani, 2008). Along these lines, astrocytes are the predominant cell type within the neurovascular unit, providing trophic support for neurons, regulating cerebral blood flow, and maintaining ionic and neurotransmitter homeostasis under physiological conditions. Conversely, astrocytes may generate cerebral innate immune responses after injury or infection, releasing pro-inflammatory mediators (Farina, et al., 2007).

AQP4, a bidirectional water channel expressed in the perivascular end feet of astrocytes, mediated glial swelling in vitro and was associated with the development of cellular edema after TBI in humans and rodents (Badaut, et al., 2011; Hu, et al., 2005). Although causative studies remain unperformed after neurotrauma, attenuated swelling of pericapillary astrocytic foot processes, decreased cellular edema, and reduced mortality were observed in AQP4-deficient mice after ischemic stroke or acute water intoxication (Manley, et al., 2000). Additionally, genetic deletion of AQP4 attenuated astrocytic migration and glial scar formation, implicating AQP4 as a potential therapeutic target to restrict deleterious astrocytic responses to injury (Saadoun, et al., 2005). Unfortunately, clinically-efficacious drugs to inhibit AQP4 expression/function do not currently exist, at least in part, due to the limited understanding of AQP4 regulation at the cellular level. Notably, we and others recently identified IL-1β as a positive regulator of AQP4 expression in cultured astrocytes and in the mouse cerebral cortex (H. Ito, et al., 2006; Laird, et al., 2010). IL-1β expression is rapidly increased following brain insults and functionally promotes reactive astrogliosis after penetrating brain injury (Lin, et al., 2006). Furthermore, elevated concentrations of IL-1β in the CSF of head trauma patients correlated with an unfavorable clinical outcome (Chiaretti, et al., 2005; Hayakata, et al., 2004). Based on these findings, we hypothesized

that strategies which reduce post-traumatic IL-1 β may effectively limit neurovascular injury after TBI.

IL-1β is synthesized as a biologically inactive 31-kDa precursor protein that requires proteolytic cleavage to generate the mature, biologically-active 17.5 kDa protein (Perregaux & Gabel, 1998). Expression of caspase-1 (also called interleukin-1 converting enzyme; ICE), the principal enzyme involved in the processing of pro-IL-1\beta into the mature IL-1\beta form, was upregulated within the rat forebrain after fluid percussion injury (Yakovlev, et al., 1997). Activated caspase-1 was strongly increased in brain tissue resected from both pediatric and adult TBI patients whereas pro-caspase-1 exhibited a decrease in expression as compared to control patients (Clark, et al., 1999; Satchell, et al., 2005). Furthermore, activated caspase-1 was elevated within the CSF of pediatric TBI patients, an observation that directly correlated with a concomitant increase in IL-1\beta and reduction in pro-IL-1\beta in these same patients (Satchell, et al., 2005). Functionally, genetic or pharmacological inhibition of caspase-1 reduced secondary tissue damage after experimental TBI in mice (Fink, et al., 1999). Taken together, these findings suggest clinical significance for caspase-1 activation after TBI and imply therapeutic targeting of caspase-1 pathway may improve outcomes.

The precise cellular mechanisms underlying caspase-1 activation remain poorly defined; however, repetitive or prolonged exposure to high concentrations of ATP increased the activation and the externalization of caspase-1 and promoted the formation of a large membrane pore required for the extracellular release of IL-1β (Laliberte, Eggler, & Gabel, 1999; Mariathasan, et al., 2006). ATP, an intracellular energy source under physiological conditions, is rapidly released into the extracellular space after traumatic or ischemic

injuries (Khakh & North, 2006; Latini, Corsi, Pedata, & Pepeu, 1996; Peng, et al., 2009; Ralevic & Burnstock, 1998). Although the functional significance remains poorly defined, the release of extracellular ATP promoted secondary tissue damage after traumatic spinal cord injury (Peng, et al., 2009). Furthermore, elevated levels of ATP metabolites within the CSF of a head trauma patient correlated with edemic development and elevated ICP (Cristofori, et al., 2005), implying a detrimental role for purinergic signaling after neurological injury.

The biological actions of ATP are mediated, at least in part, by activation of either metabotropic P2Y receptors or ionotropic P2X receptors (Ralevic & Burnstock, 1998). Among the purine receptor family members, P2X7 is a low-affinity receptor that preferentially responds to sustained elevations in ATP such as those which occurs after trauma, suggesting P2X7 possesses the optimal biophysical properties for mediating the detrimental actions of ATP after a brain injury. Herein, P2X7 specifically co-localized within astrocytic end feet within the brain, directly overlapping with the expression of AQP4. Consistent with a report showing extracellular ATP induced stellation and increased GFAP expression in astrocyte cultures (Neary, Baker, Jorgensen, & Norenberg, 1994), clinically-achievable doses of BBG decreased IL-1β production, reduced astrocytic activation, as assessed by GFAP expression, attenuated AQP4 expression, and limited cerebral edema after TBI in mice. Given the importance of cerebral edema and elevated ICP in patient mortality and long-term morbidity after TBI, P2X7 antagonism may improve acute clinical outcomes following TBI.

Increased rates of depression, aggression, and anxiety are observed over the first year in up to 51% of TBI survivors (Fann, et al., 2004); yet, a recent meta-analysis of 223 preclinical trials failed to identify any single intervention that significantly improved these

neurological outcomes after TBI (Wheaton, Mathias, & Vink, 2011). Interestingly, patients with idiopathic intracranial hypertension, a neurological disorder characterized by nontraumatic elevations in ICP, exhibited higher rates of developing depression and anxiety, as compared to matched control patients (Kleinschmidt, Digre, & Hanover, 2000). These clinical findings suggested post-traumatic elevations in ICP could directly induce psychiatric co-morbidities. IL-1β, which clinically correlates with elevated ICP after TBI (Chiaretti, et al., 2005; Holmin & Hojeberg, 2004; Shiozaki, et al., 2005), is implicated in the pathophysiology of depressive and anxiety (Koo & Duman, 2008, 2009a, 2009b; Norman, et al., 2010). Thus, the production of IL-1\beta may provide a key mechanistic bridge between acute traumatic injury and long-term neurological outcomes. Consistent with this notion, post-injury administration of clinically-relevant doses of BBG that reduced IL-1β expression and limited post-traumatic edema significantly attenuated the manifestation of depressive-like and anxious behavior after TBI. This finding is in line with a report showing P2X7-/- mice exhibited an anti-depressive-like profile and increased responsiveness to antidepressant drugs under basal conditions, as compared to wild-type mice (Basso, et al., 2009). The novel findings presented herein provide support for the notion that acute neuroinflammatory mediators contribute to elevations in ICP as well as influence the development of subsequent neurobehavioral outcomes after TBI.

Several caveats of this study warrant further consideration. Although considered a highly selective P2X7 antagonist, BBG also can inhibit both P2X2 and P2X5, albeit less potently than at P2X7 (Jiang, et al., 2000). Despite our data showing P2X7-/- mice exhibit similar responses to BBG-treated mice, we cannot exclude the possibility that off-target effects on receptors other than P2X7 mediated the beneficial actions of BBG. Similarly, it

remains unclear whether BBG penetrates the blood-brain barrier. We observed a significant accumulation of BBG within the tissue adjacent to the contusion, suggesting BBG could possibly act at the level of the CNS. Nonetheless, we cannot eliminate the possibility that BBG may also act on peripheral immune cells that express P2X7, produce pro-inflammatory mediators, and infiltrate into brain tissue after TBI. Future work by our group using cell-type specific knockout of P2X7 (e.g. astrocyte-specific P2X7 knockout) will attempt to address this issue in detail.

In conclusion, this data suggests a novel, causative role for the low-affinity ATP receptor, P2X7, in the development of cerebral edema and neurological injury after TBI. These findings also identify BBG, a drug that is well-tolerated in humans, in the treatment of cerebral edema and neurological deterioration following TBI using a clinically-feasible therapeutic window. Given the dearth of medical treatment options to limit elevated ICP and reduce co-morbid psychiatric deficits following head trauma, further exploration of P2X7 may be warranted.

References Cited

- Aarabi, B., Hesdorffer, D. C., Ahn, E. S., Aresco, C., Scalea, T. M., & Eisenberg, H. M. (2006). Outcome following decompressive craniectomy for malignant swelling due to severe head injury. *J Neurosurg*, 104(4), 469-479.
- Abbott, N. J., Revest, P. A., & Romero, I. A. (1992). Astrocyte-endothelial interaction: physiology and pathology. *Neuropathol Appl Neurobiol, 18*(5), 424-433.
- Allan, S. M., & Rothwell, N. J. (2001). Cytokines and acute neurodegeneration. *Nat Rev Neurosci, 2*(10), 734-744.
- Allen, C. H., & Ward, J. D. (1998). An evidence-based approach to management of increased intracranial pressure. *Crit Care Clin, 14*(3), 485-495.
- Amaducci, L., Forno, K. I., & Eng, L. F. (1981). Glial fibrillary acidic protein in cryogenic lesions of the rat brain. *Neurosci Lett, 21*(1), 27-32.
- Amiry-Moghaddam, M., Otsuka, T., Hurn, P. D., Traystman, R. J., Haug, F. M., Froehner, S. C., et al. (2003). An alpha-syntrophin-dependent pool of AQP4 in astroglial end-feet confers bidirectional water flow between blood and brain. *Proc Natl Acad Sci U S A, 100*(4), 2106-2111.
- Amiry-Moghaddam, M., Xue, R., Haug, F. M., Neely, J. D., Bhardwaj, A., Agre, P., et al. (2004). Alpha-syntrophin deletion removes the perivascular but not endothelial pool of aquaporin-4 at the blood-brain barrier and delays the development of brain edema in an experimental model of acute hyponatremia. *Faseb J, 18*(3), 542-544.
- Amorini, A. M., Dunbar, J. G., & Marmarou, A. (2003). Modulation of aquaporin-4 water transport in a model of TBI. *Acta Neurochir Suppl, 86*, 261-263.
- Ankarcrona, M., Dypbukt, J. M., Bonfoco, E., Zhivotovsky, B., Orrenius, S., Lipton, S. A., et al. (1995). Glutamate-induced neuronal death: a succession of necrosis or apoptosis depending on mitochondrial function. *Neuron*, *15*(4), 961-973.
- Aoki-Yoshino, K., Uchihara, T., Duyckaerts, C., Nakamura, A., Hauw, J. J., & Wakayama, Y. (2005). Enhanced expression of aquaporin 4 in human brain with inflammatory diseases. *Acta Neuropathol, 110*(3), 281-288.
- Arbeloa, J., Perez-Samartin, A., Gottlieb, M., & Matute, C. (2011). P2X7 receptor blockade prevents ATP excitotoxicity in neurons and reduces brain damage after ischemia. *Neurobiol Dis*.
- Badaut, J., Ashwal, S., Adami, A., Tone, B., Recker, R., Spagnoli, D., et al. (2011). Brain water mobility decreases after astrocytic aquaporin-4 inhibition using RNA interference. *J Cereb Blood Flow Metab, 31*(3), 819-831.
- Badaut, J., Lasbennes, F., Magistretti, P. J., & Regli, L. (2002). Aquaporins in brain: distribution, physiology, and pathophysiology. *J Cereb Blood Flow Metab*, *22*(4), 367-378.
- Bae, M. K., Kim, S. R., Lee, H. J., Wee, H. J., Yoo, M. A., Ock Oh, S., et al. (2006). Aspirin-induced blockade of NF-kappaB activity restrains up-regulation of glial fibrillary acidic protein in human astroglial cells. *Biochim Biophys Acta, 1763*(3), 282-289.
- Baldwin, S. A., & Scheff, S. W. (1996). Intermediate filament change in astrocytes following mild cortical contusion. *Glia*, *16*(3), 266-275.
- Basso, A. M., Bratcher, N. A., Harris, R. R., Jarvis, M. F., Decker, M. W., & Rueter, L. E. (2009). Behavioral profile of P2X7 receptor knockout mice in animal models of

- depression and anxiety: relevance for neuropsychiatric disorders. *Behav Brain Res, 198*(1), 83-90.
- Bignami, A. (1991). Glial cells in the central nervous system. In M. P.J. (Ed.), *Discussions in neuroscience* (Vol. VIII, pp. 1-45). Amsterdam: Elsevier.
- Binder, S., Corrigan, J. D., & Langlois, J. A. (2005). The public health approach to traumatic brain injury: an overview of CDC's research and programs. *J Head Trauma Rehabil*, 20(3), 189-195.
- Bonfoco, E., Krainc, D., Ankarcrona, M., Nicotera, P., & Lipton, S. A. (1995). Apoptosis and necrosis: two distinct events induced, respectively, by mild and intense insults with N-methyl-D-aspartate or nitric oxide/superoxide in cortical cell cultures. *Proc Natl Acad Sci U S A*, *92*(16), 7162-7166.
- Borgnia, M., Nielsen, S., Engel, A., & Agre, P. (1999). Cellular and molecular biology of the aquaporin water channels. *Annu Rev Biochem, 68*, 425-458.
- Bradshaw, B. D. (2008). Report to the Surgeon General Traumatic Brain Injury Task Force Retrieved 6 Jan 2010, 2010, from http://www.armymedicine.army.mil/reports/tbi/TBITaskForceReportJanuary2008.pdf
- The Brain Trauma Foundation. The American Association of Neurological Surgeons. The Joint Section on Neurotrauma and Critical Care. Use of barbiturates in the control of intracranial hypertension (2000). *J Neurotrauma*, 17(6-7), 527-530.
- Bramlett, H. M., & Dietrich, W. D. (2004). Pathophysiology of cerebral ischemia and brain trauma: similarities and differences. *J Cereb Blood Flow Metab, 24*(2), 133-150.
- Bratton, S. L., Chestnut, R. M., Ghajar, J., McConnell Hammond, F. F., Harris, O. A., Hartl, R., et al. (2007). Guidelines for the management of severe traumatic brain injury. VI. Indications for intracranial pressure monitoring. *J Neurotrauma, 24 Suppl 1*, S37-44.
- Bruce, D. A., Langfitt, T. W., Miller, J. D., Schutz, H., Vapalahti, M. P., Stanek, A., et al. (1973). Regional cerebral blood flow, intracranial pressure, and brain metabolism in comatose patients. *J Neurosurg*, *38*(2), 131-144.
- Bullock, R., Maxwell, W. L., Graham, D. I., Teasdale, G. M., & Adams, J. H. (1991). Glial swelling following human cerebral contusion: an ultrastructural study. *J Neurol Neurosurg Psychiatry*, *54*(5), 427-434.
- Burke, A. M., Quest, D. O., Chien, S., & Cerri, C. (1981). The effects of mannitol on blood viscosity. *J Neurosurg*, *55*(4), 550-553.
- Bush, T. G., Puvanachandra, N., Horner, C. H., Polito, A., Ostenfeld, T., Svendsen, C. N., et al. (1999). Leukocyte infiltration, neuronal degeneration, and neurite outgrowth after ablation of scar-forming, reactive astrocytes in adult transgenic mice. *Neuron*, *23*(2), 297-308.
- Cankurtaran-Sayar, S., Sayar, K., & Ugur, M. (2009). P2X7 receptor activates multiple selective dye-permeation pathways in RAW 264.7 and human embryonic kidney 293 cells. *Mol Pharmacol*, 76(6), 1323-1332.
- Caso, J. R., Pradillo, J. M., Hurtado, O., Lorenzo, P., Moro, M. A., & Lizasoain, I. (2007). Toll-like receptor 4 is involved in brain damage and inflammation after experimental stroke. *Circulation*, 115(12), 1599-1608.

Castejon, O. J. (1998). Morphological astrocytic changes in complicated human brain trauma. A light and electron microscopic study. *Brain Inj, 12*(5), 409-427; discussion 407.

- Chang, E. F., Meeker, M., & Holland, M. C. (2006). Acute traumatic intraparenchymal hemorrhage: risk factors for progression in the early post-injury period. *Neurosurgery*, *58*(4), 647-656; discussion 647-656.
- Chiaretti, A., Genovese, O., Aloe, L., Antonelli, A., Piastra, M., Polidori, G., et al. (2005). Interleukin 1beta and interleukin 6 relationship with paediatric head trauma severity and outcome. *Childs Nerv Syst, 21*(3), 185-193; discussion 194.
- Clark, R. S., Kochanek, P. M., Chen, M., Watkins, S. C., Marion, D. W., Chen, J., et al. (1999). Increases in Bcl-2 and cleavage of caspase-1 and caspase-3 in human brain after head injury. *Faseb J, 13*(8), 813-821.
- Clausen, F., Hanell, A., Bjork, M., Hillered, L., Mir, A. K., Gram, H., et al. (2009). Neutralization of interleukin-1beta modifies the inflammatory response and improves histological and cognitive outcome following traumatic brain injury in mice. *Eur J Neurosci, 30*(3), 385-396.
- Clausen, F., Hanell, A., Israelsson, C., Hedin, J., Ebendal, T., Mir, A. K., et al. (2011). Neutralization of interleukin-1beta reduces cerebral edema and tissue loss and improves late cognitive outcome following traumatic brain injury in mice. *Eur J Neurosci*, *34*(1), 110-123.
- Coddou, C., Yan, Z., Obsil, T., Huidobro-Toro, J. P., & Stojilkovic, S. S. (2011). Activation and regulation of purinergic P2X receptor channels. *Pharmacol Rev, 63*(3), 641-683.
- Cohen, J. E., Rajz, G., Itshayek, E., & Umansky, F. (2004). Bilateral acute epidural hematoma after evacuation of acute subdural hematoma: brain shift and the dynamics of extraoxial collections. *Neurol Res*, *26*(7), 763-766.
- Cold, G. E., & Jensen, F. T. (1978). Cerebral autoregulation in unconscious patients with brain injury. *Acta Anaesthesiol Scand*, *22*(3), 270-280.
- Cold, G. E., & Jensen, F. T. (1980). Cerebral blood flow in the acute phase after head injury. Part 1: Correlation to age of the patients, clinical outcome and localisation of the injured region. *Acta Anaesthesiol Scand*, 24(3), 245-251.
- Collo, G., Neidhart, S., Kawashima, E., Kosco-Vilbois, M., North, R. A., & Buell, G. (1997). Tissue distribution of the P2X7 receptor. *Neuropharmacology*, *36*(9), 1277-1283.
- Cormio, M., Gopinath, S. P., Valadka, A., & Robertson, C. S. (1999). Cerebral hemodynamic effects of pentobarbital coma in head-injured patients. J *Neurotrauma*, 16(10), 927-936.
- Cortez, S. C., McIntosh, T. K., & Noble, L. J. (1989). Experimental fluid percussion brain injury: vascular disruption and neuronal and glial alterations. *Brain Res, 482*(2), 271-282.
- Cotrina, M. L., & Nedergaard, M. (2009). Physiological and pathological functions of P2X7 receptor in the spinal cord. *Purinergic Signal*, *5*(2), 223-232.
- Cremer, O. L., van Dijk, G. W., van Wensen, E., Brekelmans, G. J., Moons, K. G., Leenen, L. P., et al. (2005). Effect of intracranial pressure monitoring and targeted intensive care on functional outcome after severe head injury. *Crit Care Med, 33*(10), 2207-2213.

Cristofori, L., Tavazzi, B., Gambin, R., Vagnozzi, R., Signoretti, S., Amorini, A. M., et al. (2005). Biochemical analysis of the cerebrospinal fluid: evidence for catastrophic energy failure and oxidative damage preceding brain death in severe head injury: a case report. *Clin Biochem, 38*(1), 97-100.

- Cruz, J., Miner, M. E., Allen, S. J., Alves, W. M., & Gennarelli, T. A. (1990). Continuous monitoring of cerebral oxygenation in acute brain injury: injection of mannitol during hyperventilation. *J Neurosurg*, *73*(5), 725-730.
- Davis, A. E. (2000). Mechanisms of traumatic brain injury: biomechanical, structural and cellular considerations. *Crit Care Nurs Q, 23*(3), 1-13.
- De Bonis, P., Pompucci, A., Mangiola, A., Paternoster, G., Festa, R., Nucci, C. G., et al. (2011). Decompressive Craniectomy for Elderly Patients with Traumatic Brain Injury: it's Probably not Worth the While. *J Neurotrauma*.
- Dempsey, R. J., Baskaya, M. K., & Dogan, A. (2000). Attenuation of brain edema, blood-brain barrier breakdown, and injury volume by ifenprodil, a polyamine-site N-methyl-D-aspartate receptor antagonist, after experimental traumatic brain injury in rats. *Neurosurgery*, 47(2), 399-404; discussion 404-396.
- Deuchars, S. A., Atkinson, L., Brooke, R. E., Musa, H., Milligan, C. J., Batten, T. F., et al. (2001). Neuronal P2X7 receptors are targeted to presynaptic terminals in the central and peripheral nervous systems. *J Neurosci, 21*(18), 7143-7152.
- Dhandapani, K., & Brann, D. (2003). Neuroprotective effects of estrogen and tamoxifen in vitro: a facilitative role for glia? *Endocrine*, *21*(1), 59-66.
- Dhandapani, K. M., Hadman, M., De Sevilla, L., Wade, M. F., Mahesh, V. B., & Brann, D. W. (2003). Astrocyte protection of neurons: role of transforming growth factorbeta signaling via a c-Jun-AP-1 protective pathway. *J Biol Chem, 278*(44), 43329-43339.
- Dhandapani, K. M., Wade, F. M., Mahesh, V. B., & Brann, D. W. (2005). Astrocyte-derived transforming growth factor-{beta} mediates the neuroprotective effects of 17{beta}-estradiol: involvement of nonclassical genomic signaling pathways. *Endocrinology*, 146(6), 2749-2759.
- Dietrich, W. D., Truettner, J., Zhao, W., Alonso, O. F., Busto, R., & Ginsberg, M. D. (1999). Sequential changes in glial fibrillary acidic protein and gene expression following parasagittal fluid-percussion brain injury in rats. *J Neurotrauma*, *16*(7), 567-581.
- Ding, J. Y., Kreipke, C. W., Speirs, S. L., Schafer, P., Schafer, S., & Rafols, J. A. (2009). Hypoxia-inducible factor-1alpha signaling in aquaporin upregulation after traumatic brain injury. *Neurosci Lett*, *453*(1), 68-72.
- Donato, T., Shapira, Y., Artru, A., & Powers, K. (1994). Effect of mannitol on cerebrospinal fluid dynamics and brain tissue edema. *Anesth Analg, 78*(1), 58-66.
- Eddleston, M., & Mucke, L. (1993). Molecular profile of reactive astrocytes--implications for their role in neurologic disease. *Neuroscience*, *54*(1), 15-36.
- Eisenberg, H. M., Gary, H. E., Jr., Aldrich, E. F., Saydjari, C., Turner, B., Foulkes, M. A., et al. (1990). Initial CT findings in 753 patients with severe head injury. A report from the NIH Traumatic Coma Data Bank. *J Neurosurg*, 73(5), 688-698.
- Enevoldsen, E. M., & Jensen, F. T. (1978). Autoregulation and CO2 responses of cerebral blood flow in patients with acute severe head injury. *J Neurosurg, 48*(5), 689-703.

Faden, A. I., Demediuk, P., Panter, S. S., & Vink, R. (1989). The role of excitatory amino acids and NMDA receptors in traumatic brain injury. *Science*, *244*(4906), 798-800.

- Faden, A. I., O'Leary, D. M., Fan, L., Bao, W., Mullins, P. G., & Movsesyan, V. A. (2001). Selective blockade of the mGluR1 receptor reduces traumatic neuronal injury in vitro and improvesoOutcome after brain trauma. *Exp Neurol*, 167(2), 435-444.
- Fan, L., Young, P. R., Barone, F. C., Feuerstein, G. Z., Smith, D. H., & McIntosh, T. K. (1995). Experimental brain injury induces expression of interleukin-1 beta mRNA in the rat brain. *Brain Res Mol Brain Res*, *30*(1), 125-130.
- Fann, J. R., Burington, B., Leonetti, A., Jaffe, K., Katon, W. J., & Thompson, R. S. (2004). Psychiatric illness following traumatic brain injury in an adult health maintenance organization population. *Arch Gen Psychiatry*, *61*(1), 53-61.
- Farina, C., Aloisi, F., & Meinl, E. (2007). Astrocytes are active players in cerebral innate immunity. *Trends Immunol, 28*(3), 138-145.
- Faul M, X. L., Wald MM, Coronado VG (2010). Traumatic Brain Injury in the United States: Emergency Department Visits, Hospitalizaions and Deaths 2002-2006
- Faul, M., Xu, L., Wald, M. M., & Coronado, V. G. (Eds.). (2010). *Traumatic Brain Injury in the United States: Emergency Department visits, hospitalizations and deaths 2002-2006.* Atlanta, GA: Centers for Disease Control and Prevention, National Center for Injury Prevention and Control.
- Ferrari, D., Pizzirani, C., Adinolfi, E., Lemoli, R. M., Curti, A., Idzko, M., et al. (2006). The P2X7 receptor: a key player in IL-1 processing and release. *J Immunol*, 176(7), 3877-3883.
- Fink, K. B., Andrews, L. J., Butler, W. E., Ona, V. O., Li, M., Bogdanov, M., et al. (1999). Reduction of post-traumatic brain injury and free radical production by inhibition of the caspase-1 cascade. *Neuroscience*, *94*(4), 1213-1218.
- Gaetz, M. (2004). The neurophysiology of brain injury. *Clin Neurophysiol*, 115(1), 4-18.
- Ghabriel, M. N., Thomas, A., & Vink, R. (2006). Magnesium restores altered aquaporin-4 immunoreactivity following traumatic brain injury to a pre-injury state. *Acta Neurochir Suppl*, *96*, 402-406.
- Gooch, M. R., Gin, G. E., Kenning, T. J., & German, J. W. (2009). Complications of cranioplasty following decompressive craniectomy: analysis of 62 cases. *Neurosurg Focus*, *26*(6), E9.
- Grant, F. C., & Norcross, N. C. (1939). Repair of Cranial Defects by Cranioplasty. *Ann Surg*, 110(4), 488-512.
- Grant, G. A., Jolley, M., Ellenbogen, R. G., Roberts, T. S., Gruss, J. R., & Loeser, J. D. (2004). Failure of autologous bone-assisted cranioplasty following decompressive craniectomy in children and adolescents. *J Neurosurg*, 100(2 Suppl Pediatrics), 163-168.
- Guerra, W. K., Gaab, M. R., Dietz, H., Mueller, J. U., Piek, J., & Fritsch, M. J. (1999). Surgical decompression for traumatic brain swelling: indications and results. *J Neurosurg*, 90(2), 187-196.
- Guo, Q., Sayeed, I., Baronne, L. M., Hoffman, S. W., Guennoun, R., & Stein, D. G. (2006). Progesterone administration modulates AQP4 expression and edema after traumatic brain injury in male rats. *Exp Neurol*, 198(2), 469-478.

Hanamsagar, R., Torres, V., & Kielian, T. Inflammasome activation and IL-1beta/IL-18 processing are influenced by distinct pathways in microglia. *J Neurochem*, 119(4), 736-748.

- Hayakata, T., Shiozaki, T., Tasaki, O., Ikegawa, H., Inoue, Y., Toshiyuki, F., et al. (2004). Changes in CSF S100B and cytokine concentrations in early-phase severe traumatic brain injury. *Shock*, *22*(2), 102-107.
- Hayashi, Y., Nomura, M., Yamagishi, S., Harada, S., Yamashita, J., & Yamamoto, H. (1997). Induction of various blood-brain barrier properties in non-neural endothelial cells by close apposition to co-cultured astrocytes. *Glia*, 19(1), 13-26.
- Hinkle, D. A., Baldwin, S. A., Scheff, S. W., & Wise, P. M. (1997). GFAP and S100beta expression in the cortex and hippocampus in response to mild cortical contusion. *J Neurotrauma*, *14*(10), 729-738.
- Ho, K. M., Honeybul, S., & Litton, E. (2011). Delayed neurological recovery after decompressive craniectomy for severe nonpenetrating traumatic brain injury. *Crit Care Med*.
- Holmin, S., & Hojeberg, B. (2004). In situ detection of intracerebral cytokine expression after human brain contusion. *Neurosci Lett, 369*(2), 108-114.
- Honeybul, S. (2010). Complications of decompressive craniectomy for head injury. J *Clin Neurosci*, 17(4), 430-435.
- Honeybul, S., & Ho, K. M. (2011). Long-term complications of decompressive craniectomy for head injury. *J Neurotrauma*, *28*(6), 929-935.
- Howard, J. L., Cipolle, M. D., Anderson, M., Sabella, V., Shollenberger, D., Li, P. M., et al. (2008). Outcome after decompressive craniectomy for the treatment of severe traumatic brain injury. *J Trauma*, *65*(2), 380-385; discussion 385-386.
- Hoyte, L., Barber, P. A., Buchan, A. M., & Hill, M. D. (2004). The rise and fall of NMDA antagonists for ischemic stroke. *Curr Mol Med*, *4*(2), 131-136.
- Hu, H., Yao, H. T., Zhang, W. P., Zhang, L., Ding, W., Zhang, S. H., et al. (2005). Increased expression of aquaporin-4 in human traumatic brain injury and brain tumors. *J Zhejiang Univ Sci B, 6*(1), 33-37.
- Hutchinson, P. J., O'Connell, M. T., Rothwell, N. J., Hopkins, S. J., Nortje, J., Carpenter, K. L., et al. (2007). Inflammation in human brain injury: intracerebral concentrations of IL-1alpha, IL-1beta, and their endogenous inhibitor IL-1ra. *J Neurotrauma*, *24*(10), 1545-1557.
- Iadecola, C. (2004). Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci*, *5*(5), 347-360.
- Ikonomidou, C., & Turski, L. (2002). Why did NMDA receptor antagonists fail clinical trials for stroke and traumatic brain injury? *Lancet Neurol, 1*(6), 383-386.
- Ito, H., Yamamoto, N., Arima, H., Hirate, H., Morishima, T., Umenishi, F., et al. (2006). Interleukin-1beta induces the expression of aquaporin-4 through a nuclear factor-kappaB pathway in rat astrocytes. *J Neurochem, 99*(1), 107-118.
- Ito, J., Marmarou, A., Barzo, P., Fatouros, P., & Corwin, F. (1996). Characterization of edema by diffusion-weighted imaging in experimental traumatic brain injury. J Neurosurg, 84(1), 97-103.
- Jafar, J. J., Johns, L. M., & Mullan, S. F. (1986). The effect of mannitol on cerebral blood flow. *J Neurosurg*, *64*(5), 754-759.
- Janzer, R. C., & Raff, M. C. (1987). Astrocytes induce blood-brain barrier properties in endothelial cells. *Nature*, *325*(6101), 253-257.

Jiang, L. H., Mackenzie, A. B., North, R. A., & Surprenant, A. (2000). Brilliant blue G selectively blocks ATP-gated rat P2X(7) receptors. *Mol Pharmacol*, *58*(1), 82-88.

- Kan, P., Amini, A., Hansen, K., White, G. L., Jr., Brockmeyer, D. L., Walker, M. L., et al. (2006). Outcomes after decompressive craniectomy for severe traumatic brain injury in children. *J Neurosurg*, 105(5 Suppl), 337-342.
- Katayama, Y., Tsubokawa, T., Miyazaki, S., Kawamata, T., & Yoshino, A. (1990). Oedema fluid formation within contused brain tissue as a cause of medically uncontrollable elevation of intracranial pressure: the role of surgical therapy. *Acta Neurochir Suppl (Wien), 51*, 308-310.
- Kaufmann, A. M., & Cardoso, E. R. (1992). Aggravation of vasogenic cerebral edema by multiple-dose mannitol. *J Neurosurg*, 77(4), 584-589.
- Kawamata, T., & Katayama, Y. (2006). Surgical management of early massive edema caused by cerebral contusion in head trauma patients. *Acta Neurochir Suppl, 96*, 3-6.
- Kawamata, T., & Katayama, Y. (2007). Cerebral contusion: a role model for lesion progression. *Prog Brain Res, 161*, 235-241.
- Khakh, B. S., & North, R. A. (2006). P2X receptors as cell-surface ATP sensors in health and disease. *Nature*, 442(7102), 527-532.
- Kiening, K. L., van Landeghem, F. K., Schreiber, S., Thomale, U. W., von Deimling, A., Unterberg, A. W., et al. (2002). Decreased hemispheric Aquaporin-4 is linked to evolving brain edema following controlled cortical impact injury in rats. *Neurosci Lett*, 324(2), 105-108.
- Kimbler, D. E., Murphy, M., & Dhandapani, K. M. (2011). Concussion and the adolescent athlete. *J Neurosci Nurs*, *43*(6), 286-290.
- King, M. D., McCracken, D. J., Wade, F. M., Meiler, S. E., Alleyne, C. H., & Dhandapani, K. M. (2011). Attenuation of hematoma size and neurological injury with curcumin following intracerebral hemorrhage in mice. *J Neurosurg, 115*(1), 116-123.
- Kinoshita, K., Chatzipanteli, K., Vitarbo, E., Truettner, J. S., Alonso, O. F., & Dietrich, W. D. (2002). Interleukin-1beta messenger ribonucleic acid and protein levels after fluid-percussion brain injury in rats: importance of injury severity and brain temperature. *Neurosurgery*, *51*(1), 195-203; discussion 203.
- Klatzo, I. (1967). Presidental address. Neuropathological aspects of brain edema. *J Neuropathol Exp Neurol, 26*(1), 1-14.
- Kleinschmidt, J. J., Digre, K. B., & Hanover, R. (2000). Idiopathic intracranial hypertension: relationship to depression, anxiety, and quality of life. *Neurology*, *54*(2), 319-324.
- Kokiko, O. N., Murashov, A. K., & Hoane, M. R. (2006). Administration of raloxifene reduces sensorimotor and working memory deficits following traumatic brain injury. *Behav Brain Res*, 170(2), 233-240.
- Kontopoulos, V., Foroglou, N., Patsalas, J., Magras, J., Foroglou, G., Yiannakou-Pephtoulidou, M., et al. (2002). Decompressive craniectomy for the management of patients with refractory hypertension: should it be reconsidered? *Acta Neurochir (Wien)*, 144(8), 791-796.
- Koo, J. W., & Duman, R. S. (2008). IL-1beta is an essential mediator of the antineurogenic and anhedonic effects of stress. *Proc Natl Acad Sci U S A, 105*(2), 751-756.

Koo, J. W., & Duman, R. S. (2009a). Evidence for IL-1 receptor blockade as a therapeutic strategy for the treatment of depression. *Curr Opin Investig Drugs*, 10(7), 664-671.

- Koo, J. W., & Duman, R. S. (2009b). Interleukin-1 receptor null mutant mice show decreased anxiety-like behavior and enhanced fear memory. *Neurosci Lett, 456*(1), 39-43.
- Kotwica, Z., & Persson, L. (1991). Effect of mannitol on intracranial pressure in focal cerebral ischemia. An experimental study in a rat. *Mater Med Pol, 23*(4), 280-284.
- Lai, Y., Stange, C., Wisniewski, S. R., Adelson, P. D., Janesko-Feldman, K. L., Brown, D. S., et al. (2006). Mitochondrial heat shock protein 60 is increased in cerebrospinal fluid following pediatric traumatic brain injury. *Dev Neurosci*, 28(4-5), 336-341.
- Laird, M. D., Sukumari-Ramesh, S., Swift, A. E., Meiler, S. E., Vender, J. R., & Dhandapani, K. M. Curcumin attenuates cerebral edema following traumatic brain injury in mice: a possible role for aquaporin-4? *J Neurochem, 113*(3), 637-648.
- Laird, M. D., Sukumari-Ramesh, S., Swift, A. E., Meiler, S. E., Vender, J. R., & Dhandapani, K. M. (2010). Curcumin attenuates cerebral edema following traumatic brain injury in mice: a possible role for aquaporin-4? *J Neurochem,* 113(3), 637-648.
- Laird, M. D., Vender, J. R., & Dhandapani, K. M. (2008). Opposing roles for reactive astrocytes following traumatic brain injury. *Neurosignals*, *16*(2-3), 154-164.
- Laliberte, R. E., Eggler, J., & Gabel, C. A. (1999). ATP treatment of human monocytes promotes caspase-1 maturation and externalization. *J Biol Chem, 274*(52), 36944-36951.
- Lam, A. G., Koppal, T., Akama, K. T., Guo, L., Craft, J. M., Samy, B., et al. (2001). Mechanism of glial activation by S100B: involvement of the transcription factor NFkappaB. *Neurobiol Aging*, *22*(5), 765-772.
- Langlois, J. A., & Sattin, R. W. (2005). Traumatic brain injury in the United States: research and programs of the Centers for Disease Control and Prevention (CDC). *J Head Trauma Rehabil, 20*(3), 187-188.
- Latini, S., Corsi, C., Pedata, F., & Pepeu, G. (1995). The source of brain adenosine outflow during ischemia and electrical stimulation. *Neurochem Int, 27*(3), 239-244.
- Latini, S., Corsi, C., Pedata, F., & Pepeu, G. (1996). The source of brain adenosine outflow during ischemia and electrical stimulation. *Neurochem Int, 28*(1), 113-118.
- Lazovic, J., Basu, A., Lin, H. W., Rothstein, R. P., Krady, J. K., Smith, M. B., et al. (2005). Neuroinflammation and both cytotoxic and vasogenic edema are reduced in interleukin-1 type 1 receptor-deficient mice conferring neuroprotection. *Stroke*, *36*(10), 2226-2231.
- Le Feuvre, R., Brough, D., & Rothwell, N. (2002). Extracellular ATP and P2X7 receptors in neurodegeneration. *Eur J Pharmacol*, 447(2-3), 261-269.
- Lee, K. S., Bae, W. K., Doh, J. W., Bae, H. G., & Yun, I. G. (1998). Origin of chronic subdural haematoma and relation to traumatic subdural lesions. *Brain Inj*, 12(11), 901-910.

Lee, M. W., Deppe, S. A., Sipperly, M. E., Barrette, R. R., & Thompson, D. R. (1994). The efficacy of barbiturate coma in the management of uncontrolled intracranial hypertension following neurosurgical trauma. *J Neurotrauma*, 11(3), 325-331.

- Lee, S. C., Dickson, D. W., & Brosnan, C. F. (1995). Interleukin-1, nitric oxide and reactive astrocytes. *Brain Behav Immun*, *9*(4), 345-354.
- Lehnardt, S. (2010). Innate immunity and neuroinflammation in the CNS: the role of microglia in Toll-like receptor-mediated neuronal injury. *Glia*, *58*(3), 253-263.
- Lehnardt, S., Massillon, L., Follett, P., Jensen, F. E., Ratan, R., Rosenberg, P. A., et al. (2003). Activation of innate immunity in the CNS triggers neurodegeneration through a Toll-like receptor 4-dependent pathway. *Proc Natl Acad Sci U S A*, *100*(14), 8514-8519.
- Levin, A. B., Duff, T. A., & Javid, M. J. (1979). Treatment of increased intracranial pressure: a comparison of different hyperosmotic agents and the use of thiopental. *Neurosurgery*, *5*(5), 570-575.
- Levin, H. S., Eisenberg, H. M., Gary, H. E., Marmarou, A., Foulkes, M. A., Jane, J. A., et al. (1991). Intracranial hypertension in relation to memory functioning during the first year after severe head injury. *Neurosurgery*, *28*(2), 196-199; discussion 200.
- Lin, H. W., Basu, A., Druckman, C., Cicchese, M., Krady, J. K., & Levison, S. W. (2006). Astrogliosis is delayed in type 1 interleukin-1 receptor-null mice following a penetrating brain injury. *J Neuroinflammation*, *3*, 15.
- Liu, K., Mori, S., Takahashi, H. K., Tomono, Y., Wake, H., Kanke, T., et al. (2007). Antihigh mobility group box 1 monoclonal antibody ameliorates brain infarction induced by transient ischemia in rats. *Faseb J, 21*(14), 3904-3916.
- Lu, K. T., Wang, Y. W., Yang, J. T., Yang, Y. L., & Chen, H. I. (2005). Effect of interleukin-1 on traumatic brain injury-induced damage to hippocampal neurons. *J Neurotrauma*, *22*(8), 885-895.
- Mahesh, V. B., Dhandapani, K. M., & Brann, D. W. (2006). Role of astrocytes in reproduction and neuroprotection. *Mol Cell Endocrinol*, 246(1-2), 1-9.
- Management of Concussion/mild Traumatic Brain Injury (2009). In D. o. V. Affairs (Ed.): The Management of Concussion/mTBI Working Group.
- Manley, G. T., Fujimura, M., Ma, T., Noshita, N., Filiz, F., Bollen, A. W., et al. (2000). Aquaporin-4 deletion in mice reduces brain edema after acute water intoxication and ischemic stroke. *Nat Med*, *6*(2), 159-163.
- Manno, E. M., Adams, R. E., Derdeyn, C. P., Powers, W. J., & Diringer, M. N. (1999). The effects of mannitol on cerebral edema after large hemispheric cerebral infarct. *Neurology*, *52*(3), 583-587.
- Mariathasan, S., Weiss, D. S., Newton, K., McBride, J., O'Rourke, K., Roose-Girma, M., et al. (2006). Cryopyrin activates the inflammasome in response to toxins and ATP. *Nature*, *440*(7081), 228-232.
- Marmarou, A. (2007). A review of progress in understanding the pathophysiology and treatment of brain edema. *Neurosurg Focus, 22*(5), E1.
- Marshall, L. F., Smith, R. W., & Shapiro, H. M. (1979). The outcome with aggressive treatment in severe head injuries. Part I: the significance of intracranial pressure monitoring. *J Neurosurg*, *50*(1), 20-25.
- Matsuno, A., Katayama, H., Wada, H., Morikawa, K., Tanaka, K., Tanaka, H., et al. (2003). Significance of consecutive bilateral surgeries for patients with acute

- subdural hematoma who develop contralateral acute epi- or subdural hematoma. *Surg Neurol, 60*(1), 23-30; discussion 30.
- Miller, J. D., Becker, D. P., Ward, J. D., Sullivan, H. G., Adams, W. E., & Rosner, M. J. (1977). Significance of intracranial hypertension in severe head injury. J Neurosurg, 47(4), 503-516.
- Miller, J. D., Jones, P. A., Dearden, N. M., & Tocher, J. L. (1992). Progress in the management of head injury. *Br J Surg*, *79*(1), 60-64.
- Mingam, R., De Smedt, V., Amedee, T., Bluthe, R. M., Kelley, K. W., Dantzer, R., et al. (2008). In vitro and in vivo evidence for a role of the P2X7 receptor in the release of IL-1 beta in the murine brain. *Brain Behav Immun, 22*(2), 234-244.
- Mitka, M. (2010). Reports of concussions from youth sports rise along with awareness of the problem. *JAMA*, *304*(16), 1775-1776.
- Morgalla, M. H., Will, B. E., Roser, F., & Tatagiba, M. (2008). Do long-term results justify decompressive craniectomy after severe traumatic brain injury? *J Neurosurg*, 109(4), 685-690.
- Morganti-Kossmann, M. C., Rancan, M., Stahel, P. F., & Kossmann, T. (2002). Inflammatory response in acute traumatic brain injury: a double-edged sword. *Curr Opin Crit Care*, 8(2), 101-105.
- Muir, K. W. (2006). Glutamate-based therapeutic approaches: clinical trials with NMDA antagonists. *Curr Opin Pharmacol*, *6*(1), 53-60.
- Muizelaar, J. P., Wei, E. P., Kontos, H. A., & Becker, D. P. (1983). Mannitol causes compensatory cerebral vasoconstriction and vasodilation in response to blood viscosity changes. *J Neurosurg*, *59*(5), 822-828.
- Narayan, R. K., Kishore, P. R., Becker, D. P., Ward, J. D., Enas, G. G., Greenberg, R. P., et al. (1982). Intracranial pressure: to monitor or not to monitor? A review of our experience with severe head injury. *J Neurosurg*, *56*(5), 650-659.
- Narayan, R. K., Michel, M. E., Ansell, B., Baethmann, A., Biegon, A., Bracken, M. B., et al. (2002). Clinical trials in head injury. *J Neurotrauma*, 19(5), 503-557.
- Neary, J. T. (2000). Trophic actions of extracellular ATP: gene expression profiling by DNA array analysis. *J Auton Nerv Syst*, 81(1-3), 200-204.
- Neary, J. T., Baker, L., Jorgensen, S. L., & Norenberg, M. D. (1994). Extracellular ATP induces stellation and increases glial fibrillary acidic protein content and DNA synthesis in primary astrocyte cultures. *Acta Neuropathol, 87*(1), 8-13.
- Nicchia, G. P., Frigeri, A., Liuzzi, G. M., Santacroce, M. P., Nico, B., Procino, G., et al. (2000). Aquaporin-4-containing astrocytes sustain a temperature- and mercury-insensitive swelling in vitro. *Glia*, 31(1), 29-38.
- Nielsen, S., Nagelhus, E. A., Amiry-Moghaddam, M., Bourque, C., Agre, P., & Ottersen, O. P. (1997). Specialized membrane domains for water transport in glial cells: high-resolution immunogold cytochemistry of aquaporin-4 in rat brain. JNeurosci, 17(1), 171-180.
- Nieto-Sampedro, M., & Berman, M. A. (1987). Interleukin-1-like activity in rat brain: sources, targets, and effect of injury. *J Neurosci Res*, 17(3), 214-219.
- Node, Y., & Nakazawa, S. (1990). Clinical study of mannitol and glycerol on raised intracranial pressure and on their rebound phenomenon. *Adv Neurol*, *52*, 359-363.

Nonaka, M., Chen, X. H., Pierce, J. E., Leoni, M. J., McIntosh, T. K., Wolf, J. A., et al. (1999). Prolonged activation of NF-kappaB following traumatic brain injury in rats. *J Neurotrauma*, *16*(11), 1023-1034.

- Nordby, H. K., & Nesbakken, R. (1984). The effect of high dose barbiturate decompression after severe head injury. A controlled clinical trial. *Acta Neurochir (Wien)*, 72(3-4), 157-166.
- Nordstrom, C. H., Messeter, K., Sundbarg, G., Schalen, W., Werner, M., & Ryding, E. (1988). Cerebral blood flow, vasoreactivity, and oxygen consumption during barbiturate therapy in severe traumatic brain lesions. *J Neurosurg, 68*(3), 424-431.
- Norenberg, M. D. (1994). Astrocyte responses to CNS injury. *J Neuropathol Exp Neurol*, 53(3), 213-220.
- Norman, G. J., Karelina, K., Zhang, N., Walton, J. C., Morris, J. S., & Devries, A. C. (2010). Stress and IL-1beta contribute to the development of depressive-like behavior following peripheral nerve injury. *Mol Psychiatry*, *15*(4), 404-414.
- Nortje, J., & Menon, D. K. (2004). Traumatic brain injury: physiology, mechanisms, and outcome. *Curr Opin Neurol*, 17(6), 711-718.
- Obrenovitch, T. P., & Urenjak, J. (1997). Is high extracellular glutamate the key to excitotoxicity in traumatic brain injury? *J Neurotrauma*, *14*(10), 677-698.
- Oertel, M., Kelly, D. F., McArthur, D., Boscardin, W. J., Glenn, T. C., Lee, J. H., et al. (2002). Progressive hemorrhage after head trauma: predictors and consequences of the evolving injury. *J Neurosurg*, *96*(1), 109-116.
- Okie, S. (2005). Traumatic brain injury in the war zone. *N Engl J Med, 352*(20), 2043-2047.
- Okiyama, K., Smith, D. H., White, W. F., & McIntosh, T. K. (1998). Effects of the NMDA antagonist CP-98,113 on regional cerebral edema and cardiovascular, cognitive, and neurobehavioral function following experimental brain injury in the rat. *Brain Res*, *792*(2), 291-298.
- Ommaya, A. K., & Gennarelli, T. A. (1974). Cerebral concussion and traumatic unconsciousness. Correlation of experimental and clinical observations of blunt head injuries. *Brain*, *97*(4), 633-654.
- Overgaard, J., & Tweed, W. A. (1974). Cerebral circulation after head injury. 1. Cerebral blood flow and its regulation after closed head injury with emphasis on clinical correlations. *J Neurosurg*, *41*(5), 531-541.
- Palmer, A. M., Marion, D. W., Botscheller, M. L., Swedlow, P. E., Styren, S. D., & DeKosky, S. T. (1993). Traumatic brain injury-induced excitotoxicity assessed in a controlled cortical impact model. *J Neurochem, 61*(6), 2015-2024.
- Pan, D. S., Liu, W. G., Yang, X. F., & Cao, F. (2007). Inhibitory effect of progesterone on inflammatory factors after experimental traumatic brain injury. *Biomed Environ Sci, 20*(5), 432-438.
- Pandya, J. D., Pauly, J. R., Nukala, V. N., Sebastian, A. H., Day, K. M., Korde, A. S., et al. (2007). Post-Injury Administration of Mitochondrial Uncouplers Increases Tissue Sparing and Improves Behavioral Outcome following Traumatic Brain Injury in Rodents. *J Neurotrauma*, 24(5), 798-811.
- Panenka, W., Jijon, H., Herx, L. M., Armstrong, J. N., Feighan, D., Wei, T., et al. (2001). P2X7-like receptor activation in astrocytes increases chemokine monocyte

- chemoattractant protein-1 expression via mitogen-activated protein kinase. J *Neurosci, 21*(18), 7135-7142.
- Papadopoulos, M. C., Manley, G. T., Krishna, S., & Verkman, A. S. (2004). Aquaporin-4 facilitates reabsorption of excess fluid in vasogenic brain edema. *Faseb J, 18*(11), 1291-1293.
- Papadopoulos, M. C., & Verkman, A. S. (2007). Aquaporin-4 and brain edema. *Pediatr Nephrol*, *22*(6), 778-784.
- Pelinka, L. E., Kroepfl, A., Leixnering, M., Buchinger, W., Raabe, A., & Redl, H. (2004). GFAP versus S100B in serum after traumatic brain injury: relationship to brain damage and outcome. *J Neurotrauma*, *21*(11), 1553-1561.
- Pelinka, L. E., Kroepfl, A., Schmidhammer, R., Krenn, M., Buchinger, W., Redl, H., et al. (2004). Glial fibrillary acidic protein in serum after traumatic brain injury and multiple trauma. *J Trauma*, *57*(5), 1006-1012.
- Peng, W., Cotrina, M. L., Han, X., Yu, H., Bekar, L., Blum, L., et al. (2009). Systemic administration of an antagonist of the ATP-sensitive receptor P2X7 improves recovery after spinal cord injury. *Proc Natl Acad Sci U S A, 106*(30), 12489-12493.
- Perregaux, D. G., & Gabel, C. A. (1998). Post-translational processing of murine IL-1: evidence that ATP-induced release of IL-1 alpha and IL-1 beta occurs via a similar mechanism. *J Immunol*, *160*(5), 2469-2477.
- Pettus, E. H., Wright, D. W., Stein, D. G., & Hoffman, S. W. (2005). Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. *Brain Res, 1049*(1), 112-119.
- Pfrieger, F. W., & Barres, B. A. (1997). Synaptic efficacy enhanced by glial cells in vitro. *Science*, *277*(5332), 1684-1687.
- Phillips, L. L., Lyeth, B. G., Hamm, R. J., Reeves, T. M., & Povlishock, J. T. (1998). Glutamate antagonism during secondary deafferentation enhances cognition and axo-dendritic integrity after traumatic brain injury. *Hippocampus, 8*(4), 390-401.
- Piepmeier, J. M., & Wagner, F. C., Jr. (1982). Delayed post-traumatic extracerebral hematomas. *J Trauma*, *22*(6), 455-460.
- Pillai, A., Dhandapani, K. M., Pillai, B. A., Terry, A. V., Jr., & Mahadik, S. P. (2007). Erythropoietin Prevents Haloperidol Treatment-Induced Neuronal Apoptosis through Regulation of BDNF. *Neuropsychopharmacology*.
- Pleines, U. E., Morganti-Kossmann, M. C., Rancan, M., Joller, H., Trentz, O., & Kossmann, T. (2001). S-100 beta reflects the extent of injury and outcome, whereas neuronal specific enolase is a better indicator of neuroinflammation in patients with severe traumatic brain injury. *J Neurotrauma*, *18*(5), 491-498.
- Polin, R. S., Shaffrey, M. E., Bogaev, C. A., Tisdale, N., Germanson, T., Bocchicchio, B., et al. (1997). Decompressive bifrontal craniectomy in the treatment of severe refractory posttraumatic cerebral edema. *Neurosurgery*, *41*(1), 84-92; discussion 92-84.
- Ralevic, V., & Burnstock, G. (1998). Receptors for purines and pyrimidines. *Pharmacol Rev, 50*(3), 413-492.
- Register, F. (2006). available at http://edocket.access.gpo.gov/2006/06-3307.htm.
- Ridet, J. L., Malhotra, S. K., Privat, A., & Gage, F. H. (1997). Reactive astrocytes: cellular and molecular cues to biological function. *Trends Neurosci*, *20*(12), 570-577.

Rish, B. L., Dillon, J. D., Meirowsky, A. M., Caveness, W. F., Mohr, J. P., Kistler, J. P., et al. (1979). Cranioplasty: a review of 1030 cases of penetrating head injury. *Neurosurgery*, 4(5), 381-385.

- Roberts, I. (2000). Barbiturates for acute traumatic brain injury. *Cochrane Database Syst Rev*(2), CD000033.
- Roesler, R., Quevedo, J., & Schroder, N. (2003). Is it time to conclude that NMDA antagonists have failed? *Lancet Neurol*, *2*(1), 13; discussion 13.
- Rogers, J. M., & Read, C. A. (2007). Psychiatric comorbidity following traumatic brain injury. *Brain Inj, 21*(13-14), 1321-1333.
- Rosner, M. J. (1995). Introduction to cerebral perfusion pressure management. *Neurosurg Clin N Am, 6*(4), 761-773.
- Saadoun, S., Papadopoulos, M. C., Watanabe, H., Yan, D., Manley, G. T., & Verkman, A. S. (2005). Involvement of aquaporin-4 in astroglial cell migration and glial scar formation. *J Cell Sci, 118*(Pt 24), 5691-5698.
- Sahuquillo, J., & Arikan, F. (2006). Decompressive craniectomy for the treatment of refractory high intracranial pressure in traumatic brain injury. *Cochrane Database Syst Rev*(1), CD003983.
- Sanz, O., Acarin, L., Gonzalez, B., & Castellano, B. (2002). NF-kappaB and IkappaBalpha expression following traumatic brain injury to the immature rat brain. *J Neurosci Res, 67*(6), 772-780.
- Sarabia, R., Lobato, R. D., Rivas, J. J., Cordobes, F., Rubio, J., Cabrera, A., et al. (1988). Cerebral hemisphere swelling in severe head injury patients. *Acta Neurochir Suppl (Wien)*, 42, 40-46.
- Satchell, M. A., Lai, Y., Kochanek, P. M., Wisniewski, S. R., Fink, E. L., Siedberg, N. A., et al. (2005). Cytochrome c, a biomarker of apoptosis, is increased in cerebrospinal fluid from infants with inflicted brain injury from child abuse. *J Cereb Blood Flow Metab*, *25*(7), 919-927.
- Saul, T. G., & Ducker, T. B. (1982a). Effect of intracranial pressure monitoring and aggressive treatment on mortality in severe head injury. *J Neurosurg*, *56*(4), 498-503.
- Saul, T. G., & Ducker, T. B. (1982b). Intracranial pressure monitoring in patients with severe head injury. *Am Surg, 48*(9), 477-480.
- Schalen, W., Messeter, K., & Nordstrom, C. H. (1992). Complications and side effects during thiopentone therapy in patients with severe head injuries. *Acta Anaesthesiol Scand*, *36*(4), 369-377.
- Schwab, S., Spranger, M., Schwarz, S., & Hacke, W. (1997). Barbiturate coma in severe hemispheric stroke: useful or obsolete? *Neurology*, *48*(6), 1608-1613.
- Shiozaki, T., Hayakata, T., Tasaki, O., Hosotubo, H., Fuijita, K., Mouri, T., et al. (2005). Cerebrospinal fluid concentrations of anti-inflammatory mediators in early-phase severe traumatic brain injury. *Shock, 23*(5), 406-410.
- Singhal, A., Baker, A. J., Hare, G. M., Reinders, F. X., Schlichter, L. C., & Moulton, R. J. (2002). Association between cerebrospinal fluid interleukin-6 concentrations and outcome after severe human traumatic brain injury. *J Neurotrauma*, 19(8), 929-937.
- Sinke, A. P., Jayakumar, A. R., Panickar, K. S., Moriyama, M., Reddy, P. V., & Norenberg, M. D. (2008). NFkappaB in the mechanism of ammonia-induced astrocyte swelling in culture. *J Neurochem*, 106(6), 2302-2311.

Skaper, S. D., Debetto, P., & Giusti, P. (2010). The P2X7 purinergic receptor: from physiology to neurological disorders. *FASEB J, 24*(2), 337-345.

- Smith, D. H., Okiyama, K., Thomas, M. J., & McIntosh, T. K. (1993). Effects of the excitatory amino acid receptor antagonists kynurenate and indole-2-carboxylic acid on behavioral and neurochemical outcome following experimental brain injury. *J Neurosci*, 13(12), 5383-5392.
- Smith, J. S., Chang, E. F., Rosenthal, G., Meeker, M., von Koch, C., Manley, G. T., et al. (2007). The role of early follow-up computed tomography imaging in the management of traumatic brain injury patients with intracranial hemorrhage. *J Trauma*, *63*(1), 75-82.
- Soulet, D., & Rivest, S. (2008). Microglia. Curr Biol, 18(12), R506-508.
- Srinivasan, D., Yen, J. H., Joseph, D. J., & Friedman, W. (2004). Cell type-specific interleukin-1beta signaling in the CNS. *J Neurosci*, 24(29), 6482-6488.
- Stiver, S. I. (2009). Complications of decompressive craniectomy for traumatic brain injury. *Neurosurg Focus*, *26*(6), E7.
- Summers, C. R., Ivins, B., & Schwab, K. A. (2009). Traumatic brain injury in the United States: an epidemiologic overview. *Mt Sinai J Med*, *76*(2), 105-110.
- Sun, M. C., Honey, C. R., Berk, C., Wong, N. L., & Tsui, J. K. (2003). Regulation of aquaporin-4 in a traumatic brain injury model in rats. *J Neurosurg*, *98*(3), 565-569.
- Sviri, G. E., Aaslid, R., Douville, C. M., Moore, A., & Newell, D. W. (2009). Time course for autoregulation recovery following severe traumatic brain injury. *J Neurosurg*, 111(4), 695-700.
- Takano, T., Tian, G. F., Peng, W., Lou, N., Libionka, W., Han, X., et al. (2006). Astrocyte-mediated control of cerebral blood flow. *Nat Neurosci*, *9*(2), 260-267.
- Takenouchi, T., Sekiyama, K., Sekigawa, A., Fujita, M., Waragai, M., Sugama, S., et al. (2010). P2X7 Receptor Signaling Pathway as a Therapeutic Target for Neurodegenerative Diseases. *Arch Immunol Ther Exp (Warsz)*.
- Tan, P. G., Cincotta, M., Clavisi, O., Bragge, P., Wasiak, J., Pattuwage, L., et al. Review article: Prehospital fluid management in traumatic brain injury. *Emerg Med Australas*, *23*(6), 665-676.
- Taupin, V., Toulmond, S., Serrano, A., Benavides, J., & Zavala, F. (1993). Increase in IL-6, IL-1 and TNF levels in rat brain following traumatic lesion. Influence of preand post-traumatic treatment with Ro5 4864, a peripheral-type (p site) benzodiazepine ligand. *J Neuroimmunol, 42*(2), 177-185.
- Taya, K., Gulsen, S., Okuno, K., Prieto, R., Marmarou, C. R., & Marmarou, A. (2008). Modulation of AQP4 expression by the selective V1a receptor antagonist, SR49059, decreases trauma-induced brain edema. *Acta Neurochir Suppl, 102*, 425-429.
- Tehranian, R., Andell-Jonsson, S., Beni, S. M., Yatsiv, I., Shohami, E., Bartfai, T., et al. (2002). Improved recovery and delayed cytokine induction after closed head injury in mice with central overexpression of the secreted isoform of the interleukin-1 receptor antagonist. *J Neurotrauma*, 19(8), 939-951.
- Thiyagarajan, M., & Sharma, S. S. (2004). Neuroprotective effect of curcumin in middle cerebral artery occlusion induced focal cerebral ischemia in rats. *Life Sci, 74*(8), 969-985.

Thurman, D. J., Branche, C. M., & Sniezek, J. E. (1998). The epidemiology of sports-related traumatic brain injuries in the United States: recent developments. *J Head Trauma Rehabil*, 13(2), 1-8.

- Toulmond, S., & Rothwell, N. J. (1995). Interleukin-1 receptor antagonist inhibits neuronal damage caused by fluid percussion injury in the rat. *Brain Res, 671*(2), 261-266.
- Ullian, E. M., Christopherson, K. S., & Barres, B. A. (2004). Role for glia in synaptogenesis. *Glia*, 47(3), 209-216.
- Ullian, E. M., Sapperstein, S. K., Christopherson, K. S., & Barres, B. A. (2001). Control of synapse number by glia. *Science*, *291*(5504), 657-661.
- Unterberg, A. W., Stover, J., Kress, B., & Kiening, K. L. (2004). Edema and brain trauma. *Neuroscience*, *129*(4), 1021-1029.
- Vajda, Z., Pedersen, M., Fuchtbauer, E. M., Wertz, K., Stodkilde-Jorgensen, H., Sulyok, E., et al. (2002). Delayed onset of brain edema and mislocalization of aquaporin-4 in dystrophin-null transgenic mice. *Proc Natl Acad Sci U S A*, 99(20), 13131-13136.
- Verhoef, P. A., Estacion, M., Schilling, W., & Dubyak, G. R. (2003). P2X7 receptor-dependent blebbing and the activation of Rho-effector kinases, caspases, and IL-1 beta release. *J Immunol, 170*(11), 5728-5738.
- Verkman, A. S., & Mitra, A. K. (2000). Structure and function of aquaporin water channels. *Am J Physiol Renal Physiol, 278*(1), F13-28.
- Verkman, A. S., Yang, B., Song, Y., Manley, G. T., & Ma, T. (2000). Role of water channels in fluid transport studied by phenotype analysis of aquaporin knockout mice. *Exp Physiol, 85 Spec No*, 233S-241S.
- Verweij, B. H., Amelink, G. J., & Muizelaar, J. P. (2007). Current concepts of cerebral oxygen transport and energy metabolism after severe traumatic brain injury. *Prog Brain Res, 161*, 111-124.
- Vik, A., Nag, T., Fredriksli, O. A., Skandsen, T., Moen, K. G., Schirmer-Mikalsen, K., et al. (2008). Relationship of "dose" of intracranial hypertension to outcome in severe traumatic brain injury. *J Neurosurg*, 109(4), 678-684.
- Vitkovic, L., Bockaert, J., & Jacque, C. (2000). "Inflammatory" cytokines: neuromodulators in normal brain? *J Neurochem*, 74(2), 457-471.
- Vizuete, M. L., Venero, J. L., Vargas, C., Ilundain, A. A., Echevarria, M., Machado, A., et al. (1999). Differential upregulation of aquaporin-4 mRNA expression in reactive astrocytes after brain injury: potential role in brain edema. *Neurobiol Dis, 6*(4), 245-258.
- Wakade, C., King, M. D., Laird, M. D., Alleyne, C. H., Jr., & Dhandapani, K. M. (2009). Curcumin attenuates vascular inflammation and cerebral vasospasm after subarachnoid hemorrhage in mice. *Antioxid Redox Signal*, *11*(1), 35-45.
- Wakade, C., Sukumari-Ramesh, S., Laird, M. D., Dhandapani, K. M., & Vender, J. R. Delayed reduction in hippocampal postsynaptic density protein-95 expression temporally correlates with cognitive dysfunction following controlled cortical impact in mice. *J Neurosurg*.
- Wakade, C., Sukumari-Ramesh, S., Laird, M. D., Dhandapani, K. M., & Vender, J. R. (2010). Delayed reduction in hippocampal postsynaptic density protein-95 expression temporally correlates with cognitive dysfunction following controlled cortical impact in mice. *J Neurosurg*, 113(6), 1195-1201.

Wang, X., Arcuino, G., Takano, T., Lin, J., Peng, W. G., Wan, P., et al. (2004). P2X7 receptor inhibition improves recovery after spinal cord injury. *Nat Med, 10*(8), 821-827.

- Ward, J. D., Becker, D. P., Miller, J. D., Choi, S. C., Marmarou, A., Wood, C., et al. (1985). Failure of prophylactic barbiturate coma in the treatment of severe head injury. *J Neurosurg*, *62*(3), 383-388.
- Wheaton, P., Mathias, J. L., & Vink, R. (2011). Impact of pharmacological treatments on outcome in adult rodents after traumatic brain injury: a meta-analysis. *J Psychopharmacol*.
- Whelan-Goodinson, R., Ponsford, J., Johnston, L., & Grant, F. (2009). Psychiatric disorders following traumatic brain injury: their nature and frequency. *J Head Trauma Rehabil*, 24(5), 324-332.
- Wilson, H. L., Francis, S. E., Dower, S. K., & Crossman, D. C. (2004). Secretion of intracellular IL-1 receptor antagonist (type 1) is dependent on P2X7 receptor activation. *J Immunol*, 173(2), 1202-1208.
- Wilson, J. X. (1997). Antioxidant defense of the brain: a role for astrocytes. *Can J Physiol Pharmacol*, *75*(10-11), 1149-1163.
- Woster, P. S., & LeBlanc, K. L. (1990). Management of elevated intracranial pressure. *Clin Pharm*, *9*(10), 762-772.
- Xiang, Z., Bo, X., & Burnstock, G. (1998). Localization of ATP-gated P2X receptor immunoreactivity in rat sensory and sympathetic ganglia. *Neurosci Lett, 256*(2), 105-108.
- Yakovlev, A. G., Knoblach, S. M., Fan, L., Fox, G. B., Goodnight, R., & Faden, A. I. (1997). Activation of CPP32-like caspases contributes to neuronal apoptosis and neurological dysfunction after traumatic brain injury. *J Neurosci, 17*(19), 7415-7424.
- Yamamoto, T., Rossi, S., Stiefel, M., Doppenberg, E., Zauner, A., Bullock, R., et al. (1999). CSF and ECF glutamate concentrations in head injured patients. *Acta Neurochir Suppl, 75*, 17-19.
- Yang, X. F., Wen, L., Shen, F., Li, G., Lou, R., Liu, W. G., et al. (2008). Surgical complications secondary to decompressive craniectomy in patients with a head injury: a series of 108 consecutive cases. *Acta Neurochir (Wien)*, *150*(12), 1241-1247; discussion 1248.
- Youn, H. S., Lee, J. K., Choi, Y. J., Saitoh, S. I., Miyake, K., Hwang, D. H., et al. (2008). Cinnamaldehyde suppresses toll-like receptor 4 activation mediated through the inhibition of receptor oligomerization. *Biochem Pharmacol*, 75(2), 494-502.
- Zhao, J., Moore, A. N., Clifton, G. L., & Dash, P. K. (2005). Sulforaphane enhances aquaporin-4 expression and decreases cerebral edema following traumatic brain injury. *J Neurosci Res, 82*(4), 499-506.
- Zitnay, G. A., Zitnay, K. M., Povlishock, J. T., Hall, E. D., Marion, D. W., Trudel, T., et al. (2008). Traumatic brain injury research priorities: the Conemaugh International Brain Injury Symposium. *J Neurotrauma*, *25*(10), 1135-1152.
- Zweckberger, K., Eros, C., Zimmermann, R., Kim, S. W., Engel, D., & Plesnila, N. (2006). Effect of early and delayed decompressive craniectomy on secondary brain damage after controlled cortical impact in mice. *J Neurotrauma, 23*(7), 1083-1093.

Summary of Dissemination

Type of Dissemination	Citation	Date and Source of Approval for Public Release
Publications	Kimbler DE, Murphy M, Dhandapani KM. Concussion and the adolescent athlete J Neurosci Nurs, 43(6), 286-290.	TSNRP PAO 2011
	Kimbler DE, Shields J, Yanasak NE, Dhandapani KM. Activation of P2X7 Promotes Cerebral Edema and Neurologic Injury after Ttraumatic Brain Injury in Mice, Plos One	TSNRP PAO 2012
Poster Presentations	Kimbler DE, Dhandapani KM. Brilliant blue G (BBG), a P2X7 antagonist, reduced cerebral edema following controlled cortical impact in mice. 2010 National Neurotrauma Society Annual Meeting, Las Vegas, NV	Amedd Center and School PAO 2010
	Kimbler DE, Shields J, Dhandapani KM. P2X7 inhibition improves neurological outcomes in a murine model of traumatic brain injury. 2011 Society for Neuroscience Annual Meeting, Washington DC.	TSNRP PAO 2011

Reportable Outcomes

Reportable Outcome	Detailed Description
Applied for Patent	None
Issued a Patent	None
Developed a cell line	None
Developed a tissue or serum repository	None
Developed a data registry	None

40

Recruitment and Retention Aspect	Number
Animals Projected in Grant Application	
Animals Purchased	400
Model Development Animals	0
Research Animals	384
Animals With Complete Data	384
Animals with Incomplete Data	0

^{*}the descripency in numbers represents mice that died during procedures